J Mycol Infect 2020; 25(2): 42–44 pISSN:1226–4709, eISSN:2465–8278 http://dx.doi.org/10.17966/JMI.2020.25.2.42

JMI

Histopathological Findings of *Trichophyton rubrum* Infection in *Ex vivo* Human Skin

Hyun Ji Lee, Chihyeon Sohng, Jun Young Kim, Kyung Duck Park, Seok-Jong Lee and Weon Ju Lee^T

Department of Dermatology, School of Medicine, Kyungpook National University, Kyungpook National University Hospital, Daegu, Korea

Trichophyton rubrum (T. rubrum) is the most usual dermatophyte that is responsible for dermatophytosis in humans¹. A classic characteristic of dermatophytes is being keratinophilic. Thus, they occupy keratinized structures, including the stratum corneum of the skin, hairs, and nails. Occasionally, T. rubrum occupies the deeper skin or distant internal organ in immunosuppressed patients^{2,3}. It is unfortunate that its pathogenic mechanism of invasion into the stratum corneum and dermis is not still totally understood. Different models. such as the animal model, stripped sheets of the stratum corneum, nail plates, monolayer cell culture model, or reconstructed human epidermis, were utilized for the examination of the mechanism of dermatophyte infection⁴⁻⁸. Even though they had a few limitations to imitate dermatophyte infections in humans, they can partly exhibit the pathogenic mechanism of dermatophyte infection. In our study, we investigated the histopathological characteristics in *T. rubrum* infection with the use of ex vivo human skin. Trichophyton rubrum was cultured for 2 weeks at 24°C on Sabouraud dextrose agar. It was transferred to 10 ex vivo skin specimens which were foreskin acquired from circumcision. The ex vivo skin specimens were kept on the Sabouraud dextrose agar at 24°C for 1 week. Afterward, histopathological examination was performed using the periodic acid-Schiff-diastase (PAS-D) stain under a light microscope. In the examination, slender septate hyphae with several arthroconidia were found in the stratum corneum (Fig. 1A, 1B). Trichophyton rubrum often generates arthroconidia in vivo. Arthroconidia are involved in pathogenesis and function as a source of infection. Thicker septate hyphae and chlamydospore were found in the lower epidermis (Fig. 1B, 2A). Thicker septate hyphae and chlamydospore were also shown in the dermis (Fig. 1B, 2A, 2B). Chlamydospore is the life stage which survives in unfavorable conditions. Corzo-León et al.⁹ performed a study with *ex vivo* human skin placed on Dulbecco's Modified Eagle Medium. They exhibited long hyphae in the ex vivo skin surface 10 days following T. rubrum infection. Hyphae and arthroconidia of T. rubrum were much more demonstrated in ex vivo human skin in our study than in Corzo-León et al's. Liang et al.⁸ introduced the reconstructed human epidermis for T. rubrum infection model. In the said study, conidia and hyphae of T. rubrum were seen in the stratum corneum of the reconstructed human epidermis 4 days following its inoculation. Apart from this, infection with much more conidia of T. rubrum displayed a full epidermal invasion beyond the superficial keratinous layer. Ho et al.⁴ examined *T. rubrum* infection with the explanted porcine skin model. In the study, extended duration of infection in the skin led to luxurious growth and invasion of the dermis. The conditions might be the result of no active immune system that would restrain fungal growth, similar to our study. Invasive dermatophytoses happen more frequently in the immunocompromised host than in the healthy person. Trichophyton rubrum accounts for the majority of invasive fungal infections in immunosuppressed patients. Trichophyton rubrum in superficial dermatophytoses is confined in the stratum corneum, demon-

Received: March 11, 2020 Revised: June 8, 2020 Accepted: June 22, 2020

[†]Corresponding: Weon Ju Lee, Department of Dermatology, Kyungpook National University Hospital, 130 Dongduk-ro, Jung-gu, Daegu, 41944, Korea.

Phone: +82-53-420-5838, Fax: +82-53-426-0770, e-mail: weonju@knu.ac.kr

Copyright@2020 by The Korean Society for Medical Mycology. All right reserved.

[©]This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. http://www.ksmm.org

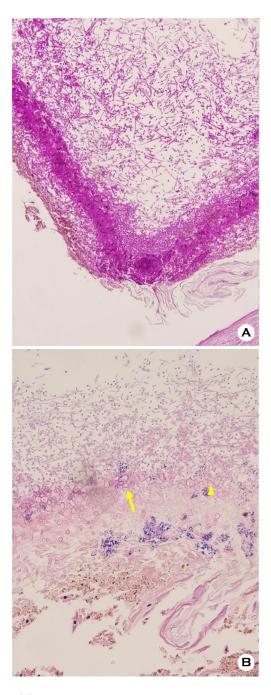


Fig. 1. (A) Slender pinkish septate hyphae with several arthroconidia in the stratum corneum (PAS-D, x200) (B) Slender pinkish septate hyphae with several arthroconidia (arrow head) in the stratum corneum and thicker septate hyphae and chlamydospore-like structures (arrow) in the lower epidermis and upper dermis (PAS-D, x400)

В

Fig. 2. (A) Thicker septate hyphae and chlamydospore-like structures (arrow) in the lower epidermis and thicker and shorter septate hyphae in the upper dermis (PAS-D, x400) (B) Thicker and shorter septate hyphae in the upper dermis (PAS-D, x400)

strating a thin septate hyphae fragment in PAS staining. Nonetheless, it is possible for *T. rubrum* to show atypical and bizarre hyphae in the dermis in invasive dermatophytoses⁵. So, these findings make suitable diagnosis hard. Our study exhibited an atypical morphology of *T. rubrum*, such as

JMI Journal of Mycology and Infection

chlamydospores and thicker hyphae, in the lower epidermis and dermis. These results will allow the recognition of the possibility of appearance changes of *T. rubrum* in invasive dermatophytoses.

Key Words: Arthroconidia, Chlamydospore, Dermatophytosis, *Ex vivo* human skin, Histopathology, *Trichophyton rubrum*

ACKNOWLEDGEMENT

There is no acknowledgment.

CONFLICT OF INTEREST

In relation to this article, we declare that there is no conflict of interest.

ORCID

Hyun Ji Lee: 0000-0002-4222-1835 Chihyeon Sohng: 0000-0002-1452-7896 Jun Young Kim: 0000-0002-2999-1018 Kyung Duck Park: 0000-0002-6067-7262 Seok-Jong Lee: 0000-0002-6131-632X Weon Ju Lee: 0000-0001-5708-1305

REFERENCES

1. Lee WJ, Kim SL, Jang YH, Lee SJ, Kim DW, Bang YJ, et al.

Increasing prevalence of *Trichophyton rubrum* identified through an analysis of 115,846 cases over the last 37 years. J Korean Med Sci 2015;30:639-643

- 2. Gong JQ, Liu XQ, Xu HB, Zeng XS, Chen W, Li XF. Deep dermatophytosis caused by *Trichophyton rubrum*: report of two cases. Mycoses 2007;50:102-108
- Squeo RF, Beer R, Silvers D, Weitzman I, Grossman M. Invasive *Trichophyton rubrum* resembling blastomycosis infection in the immunocompromised host. J Am Acad Dermatol 1998;39:379-380
- 4. Ho FK, Delgado-Charro MB, Bolhuis A. Evaluation of an explanted porcine skin model to investigate infection with the dermatophyte *Trichophyton rubrum*. Mycopathologia 2020;185:233-243
- 5. Aljabre SH, Richardson MD, Scott EM, Shankland GS. Germination of *Trichophyton mentagrophytes* on human stratum corneum *in vitro*. J Med Vet Mycol 1992;30:145 -152
- Yue X, Li Q, Wang H, Sun Y, Wang A, Zhang Q, et al. An ultrastructural study of *Trichophyton rubrum* induced onychomycosis. BMC Infect Dis 2015;15:532
- 7. Firat YH, Simanski M, Rademacher F, Schröder L, Brasch J, Harder J. Infection of keratinocytes with *Trichophytum rubrum* induces epidermal growth factor-dependent RNase 7 and human beta-defensin-3 expression. PLoS One 2014;9:e93941
- Liang PP, Huang XZ, Yi JL, Chen ZR, Ma H, Ye CX, et al. A *Trichophyton rubrum* infection model based on the reconstructed human epidermis - Episkin[®]. Chin Med J (Engl) 2016;129:54-58
- 9. Corzo-León DE, Munro CA, MacCallum DM. An *ex vivo* Human Skin Model to Study Superficial Fungal Infections. Front Microbiol 2019;10:1172